# Draft genome sequence of highly efficient arsenite-oxidizing bacterium Methylobacterium brachythecii C25, isolated from arsenic-contaminated soil

Sang Eun Jeong $^1$ , Hye Kyeong Kang $^1$ , Ji Young Jung $^2$ , Mi-Hwa Lee $^1$ , and Byung-Gon Ryu $^{1\ast}$ 

<sup>1</sup>Environmental Microbiology Research Team, Microbial Research Department, Nakdonggang National Institute of Biological Resources, Sangju 37242, Republic of Korea

<sup>2</sup> Bacterial Research Team, Microbial Research Department, Nakdonggang National Institute of Biological Resources, Sangju 37242, Republic of Korea

# 비소오염 토양 유래 세균 Methylobacterium brachythecii strain C25의 유전체

### 정상은 $^{1}$  · 강혜경 $^{1}$  · 정지영 $^{2}$  · 이미화 $^{1}$  · 류병곤 $^{1*}$

'국립낙동강생물자원관 담수생물연구본부 환경미생물연구팀, <sup>2</sup>국립낙동강생물자원관 담수생물연구본부 원핵생물연구팀

(Received December 6, 2022; Revised December 12, 2022; Accepted December 13, 2022)

Here, we report the features and draft genome sequence of Methylobacterium brachythecii C25, isolated from the agricultural soil contaminated with arsenic (As) in Republic of Korea. This strain is able to reduce the toxicity by oxidizing arsenite (As [III]) to arsenate (As [V]) while having resistance to high concentration of As (III). The genome consists of 5,191,050 bp chromosome with a GC content 66.03%, which contains 4,975 coding DNA sequences (CDSs), 49 tRNA genes, 3 rRNA genes, 4 non-coding RNA genes, and 75 pseudo genes. The genome also contains two types of As (III) oxidase (Aio, arsenite oxidase gene) and several stress resistance genes to As (III). Taken together, strain C25 is not only able oxidize As (III) to As (V) but also has resistance to As-contaminated environmental stress.

Keywords: Methylobacterium brachythecii, arsenite, biological oxidation, draft genome

Arsenic (As) as a known human carcinogen is a toxic metalloid widely present in the environment (Santini et al., 2000; Butt et al., 2011; Xiao et al., 2021). As accumulation in soil and water is mainly due to the anthropogenic activities such as use of pesticide, combustion of fossil fuels, mining and/-or smelting, and wastewater irrigation (Das et al., 2014). The predominant forms of As found in water or soil environments are inorganic As (III) (arsenite) and As (V) (arsenate) (Santini et al., 2000; Butt et al., 2011; Pous et al., 2015). As (III) is reported to be more harmful to human health than As (V) because of its higher toxicity, solubility, and mobility (Xiao et al., 2021). Microbial oxidation of As (III), which is being cheap and self-regenerating catalyst, is recently attracting considerable attention (Shi et al., 2020). In spite of the fact that understanding the mechanism or metabolism of As (III) oxidation is very important, the genetic information of the newly isolated bacterial strain, which may even suggest a novel metabolic pathway, is not yet understood (Xiao et al., 2021).

The As (III)-oxidizing bacterium Methylobacterium brachythecii C25 (KACC 19855) was isolated from the Ascontaminated agricultural soil. Contaminated samples were collected at the site of an agricultural soil in the South

<sup>\*</sup>For correspondence. E-mail: tesiakaist@gmail.com; Tel.: +82-54-530-0872; Fax: +82-54-530-0879

Chungcheong province (South Korea). The enrichment cultures were conducted aerobically at 30°C and 150 rpm for 2 weeks, using mineral salt medium (Kumar and Maitra, 2016) with 1 mM of As (III). The enrichment-cultured samples were diluted in phosphate-buffered saline (PBS) ( $pH = 7.4$ ). The diluted samples were then spread on R2A agar plates (BD Difco) and incubated at 25<sup>o</sup>C and 150 rpm for 2 weeks to obtain a single colony. To determine the oxidation capability of As (III) to As (V), each bacterial colony was seeded in 5 ml of R2A medium, and then the cells were inoculated in to the R2A medium with 1 mM of As (III) at 25°C. After 3 days, the bacterial strains that have a capability of As (III) oxidation were preferentially screened by using the silver nitrate  $(AgNO<sub>3</sub>)$ method (Dey et al., 2016; Kumari et al., 2019) and then were quantitatively measured by using the LC-ICP-MS (Akter et al., 2005). Thus, we succeed to isolate the strain C25, which had a higher oxidation capability of As (III) among the single colonies obtained. After 12 h of cultivation, 1 mM of As (III) was completely oxidized to As (V) (data not shown).

Genomic DNA was obtained from the strain C25 using Maxwell 16 DNA purification kits (Promega) according to the manufacturer's instructions. The genomic DNA was sequenced at Macrogen, Inc., using an Illumina HiSeq X Ten sequencer with 151-bp paired-end reads. The resulting 4,155,450,540 bp with 27,519,540 total reads were trimmed and assembled using SOAPdenovo. The draft genome of strain C25 consisted of 34 contigs with a total length of 5,191,050 bp (read N50, 431,687) and an average length of 152,677 bp (longest config of 1,269,555 and shortest contig of 1,094 bp). Aragon (v1.2.36) and Barrnap software (v0.6) were used to predict tRNAs and rRNAs (5S rRNAs, 16S rRNAs, and 23S rRNAs), respectively (Lagesen et al., 2007). Genome annotation was performed using the Prokaryotic Genome Annotation Pipeline (Tatusova et al., 2016). Default parameters were employed for all software unless otherwise specified.

Genome features of Methylobacterium brachythecii C25 are presented in Table 1. The assembled genome sequence coverage was 800.5x and the genomic GC content was 66.03%. A total of 4,975 coding DNA sequences (CDSs), 3 rRNA (1 each of 5S rRNA, 16S rRNA, 23S rRNA), 46 tRNA, 1 tmRNA, 4 non-coding RNA, and 75 pseudo genes were predicted. Identification of this bacterium was performed through 16S

Table 1. Genome features of Methylobacterium brachythecii C25

Genome features	Value
Genome size (bp)	5,191,050
$G + C$ content(%)	66.03
No. of rRNAs (5S, 16S, 23S)	3(1, 1, 1)
No. of tRNAs	49
No. of ncRNAs (noncoding)	$\overline{4}$
Protein coding genes	4,975
Pseudo-genes	75

rRNA gene amplification, which showed 98.72% similarity with *Methylobacterium brachythecii*  $99b<sup>T</sup>$  (Tani and Sahin, 2013).

Strain C25 has two types of arsenite oxidase (Aio) genes (locus number: HCU64\_21125, HCU64\_21130) that may lead to oxidize As (III) to As (V). The genome also contains several stress resistance genes to As (III) such as arsenite resistance transcriptional regulator (HCU64\_03160), arsenite membrane efflux pump (HCU64\_16965, HCU64\_21100, HCU64\_22845), arsenite resistance protein (HCU64\_16965, HCU64\_21705, HCU64\_22215). These defensive strategies possessed by this strain may allow it to survive in As (III)-contamination environment.

#### Nucleotide sequence accession number

The genome sequence and raw sequencing reads for strain C25 were deposited under GenBank accession number JAAUVI010000000, BioProject accession number PRJNA 615742, BioSample accession number SAMN14466051, and Sequence Read Archive (SRA) accession number SRR 22163625. Strain C25 was deposited in Korean Agricultural Culture Collection (KACC 19855).

### 전 유

중금속 오염지역으로부터 분리한 비소 산화 미생물 Methylobacterium 속 균주 C25의 유전체를 분석하였다. 이 세균은 독 성이 강한 아비산염(3가 비소, As III)를 비산염(5가 비소, As V)로 산화시키며 비소의 독성을 저감할 수 있는 호기성 미생 물이다. 유전체의 크기는 5,191,050 bp이며 G + C 함량은 66.03% 로 나타났다. 이 유전자는 4,975개의 단백질을 암호화

하는 서열을 비롯하여, 49개의 운반 RNA, 3개의 리보솜 RNA, 4개의 비번역 RNA, 75개의 유사유전자를 포함한다. 아비산 염을 비산염으로 산화시킬 수 있는 2가지 종류의 아비산염 산 화 효소(Aio, arsenite oxidase gene)를 보유하고 있다. 이러한 결과를 바탕으로, C25 균주는 아비산염을 산화하고, 아비산 염 독성에 의한 스트레스를 잘 견딜 수 있는 것으로 예측된다.

#### **Acknowledgments**

This work was carried out with support from a Nakdonggang National Institute of Biological Resources grant (project number NNIBR202202107) funded by the Ministry of Environment, South Korea.

### Conflict of Interest

The authors have no conflict of interest to report.

#### **References**

- Akter KF, Chen Z, Smith L, Davey D, and Naidu R. 2005. Speciation of arsenic in ground water samples: a comparative study of **CERACES<br>
KF, Chen Z, Smith L, Davey D, and Naidu R.** 2005. Specia<br>
of arsenic in ground water samples: a comparative stude<br>
CE-UV, HG-AAS and LC-ICP-MS. *Talanta* 68, 406–415. KF, Chen Z, Smith L, Davey D, and Naidu R. 2005. Specia<br>of arsenic in ground water samples: a comparative stud<br>CE-UV, HG-AAS and LC-ICP-MS. *Talanta* 68, 406–415.<br>S. and Rehman A. 2011. Isolation of arsenite-oxidizing bac<br>
- Butt AS and Rehman A. 2011. Isolation of arsenite-oxidizing bacteria from industrial effluents and their potential use in wastewater
- Das S, Jean JS, Kar S, Chou ML, and Chen CY. 2014. Screening of plant-growth promoting traits in arsenic-resistant bacteria isolated from agricultural soil and their potential implication for

arsenic bioremediation. J. Hazard. Mater. <sup>272</sup>, 112–120.

- Dey U, Chatterjee S, and Miondal NK. 2016. Isolation and characterization of arsenic-resistant bacteria and possible application arsenic bioremediation. *J. Hazard. Mater.* 2<br>J, Chatterjee S, and Miondal NK. 2016. Isc<br>terization of arsenic-resistant bacteria and p<br>in bioremediation. *Biotechnol. Rep.* 10, 1–7.
- Kumar V and Maitra SS. 2016. Biodegradation of endocrine disruptor V29b and the DBP degradation pathway. 3 Biotech 6, 200.
- dibutyl phthalate (DBP) by a newly isolated *Methylobacillus* sp.<br>
V29b and the DBP degradation pathway. 3 Biotech 6, 200.<br> **ri N, Rana A, and Jagadevan S.** 2019. Arsenite biotransformation<br>
by *Rhodococcus* sp.: character Kumari N, Rana A, and Jagadevan S. 2019. Arsenite biotransformation by Rhodococcus sp.: characterization, optimization using response surface methology and mechanistic studies. Sci. Total Environ. by *Rhodococcus* sp.: characterization, optimization using respons<br>surface methology and mechanistic studies. *Sci. Total Envirol*<br>687, 577–589.<br>en K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, an<br>Ussery DW. 2007. RNAm
- Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, and Ussery DW. 2007. RNAmmer: consistent and rapid annotation
- Pous N, Casentini B, Rossetti S, Fazi S, Puig S, and Aulenta F. 2015. Anaerobic arsenite oxidation with an electrode serving as the sole electron acceptor: a novel approach to the bioremediation of of ribosomal RNA genes. *Nucleic Acids Res*. **35**, 3100-3108.<br>**N, Casentini B, Rossetti S, Fazi S, Puig S, and Aulenta F. 2015.** Anaerobic arsenite oxidation with an electrode serving as the sole electron acceptor: a novel
- Santini JM, Sly LI, Schnagl RD, and Macy JM. 2000. A new chemolithotrophic arsenite oxidizing bacterium isolated from a gold mine: phylogenetic, physiological, and preliminary biochemical studies. Appl. Environ. Microbiol. Studies.<br>
Sole electron acceptor: a novel approach to the bioremediation<br> **i JM, Sly LI, Schnagl RD, and Macy JM.** 2000. A<br>
chemolithotrophic arsenite oxidizing bacterium isol
- Shi K, Wang Q, and Wang G. 2020. Microbiol. 63, 32<br>
Film K, Wang Q, and Wang G. 2020. Microbial oxidation of arsenite:<br>
regulation, chemotaxis, phosphate metabolism and energy<br>
generation. *Front. Microbiol.* 11, 569282.<br> regulation, chemotaxis, phosphate metabolism and energy generation. Front. Microbiol. 11, 569282.
- Tani A and Sahin N. 2013. Methylobacterium haplocladii sp. nov. and Methylobacterium brachythecii sp. nov., isolated from bryo-
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, and Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. Nucleic Methylobacterium brachyl<br>Methylobacterium brachyt<br>phytes. Int. J. Syst. Evol. M<br>**Daslavsky L, Lomsadze A, J. 2016.** NCBI prokaryotic *a*<br>*Acids Res.* **44**, 6614–6624.
- Xiao W, He X, Lin G, Yang Z, and Wang L. 2021. Arsenite-oxidizing bacteria isolated from an abandoned realgar minig area: characterization and the influence on arsenic accumulation in rice seedings. Environ. Technol. Innov. 12, 101800.